Maternal and newborn infants amino acid concentrations in obese women born themselves with normal and small for gestational age birth weight

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This study was undertaken to compare amino acid concentrations in maternal and newborn infants' serum in normal pregnancy and two groups of obese women who were born themselves with normal and small for gestational age (SGA) birth weight. Maternal cholesterol, lipoproteins concentrations and maternal and infants amino acid concentrations were evaluated at the time of delivery in 28 normal pregnancies, 46 obese pregnant women with normal birth weight (Ob-AGA group) and 44 obese pregnant women born themselves SGA (Ob-SGA group). Mean birth weight of newborn infants in Ob-SGA group was significantly less than in normal and Ob-AGA groups. Cholesterol and lipoproteins were significantly elevated in obese women (more prominent in Ob-SGA group). Most amino acid concentrations and fetal–maternal amino acid gradients were significantly lower in Ob-SGA group. These data suggest significant changes in placental amino acid transport/synthetic function in obese women who were born themselves SGA.

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Introduction

The prevalence of obesity among pregnant women in the world is increasing. In addition to the short-term complications of obesity during pregnancy in both mother and child, it has now been demonstrated that maternal obesity has long-term adverse effects, including childhood obesity and health risks for these children in later life.^{1,2}

The main concern is that the rising rates of childhood obesity will promote an epidemic of chronic diseases such as heart disease, arterial hypertension and 'type 2' diabetes. One view is that childhood obesity and adult diseases are being initiated by excess nutrition *in utero* or during infancy. People who become obese tend to have had a high birth weight and above average weight through infancy.^{3,4} Although fetuses of women with obesity appear to be exposed to an excess of circulating nutrients, they may not benefit as a result.

In many cases, however, early life development that leads to obesity-related disease begins with low birth weight and small body size during infancy.^{5–7}

Amino acids used both for protein synthesis and energy production represent together with glucose, fatty acids and lactate, the main nutrients during intrauterine growth.⁸

Fetal protein synthesis is dependent on both amino acid and energy supply. During positive energy balance, glucose and fatty acids utilization as fuel substitute for amino acids. In this reciprocal manner energy supply and amino acid metabolism are closely balanced.

A relationship has been demonstrated between metabolism of amino acids and apolipoproteins.⁹ Because of the incorporation into apolipoproteins synthesis, various amino acids such as leucine, glycine, valine, lysine, arginine and phenylalanine labeled with different stable isotopes have been used as tracers to study the metabolism of apolipoproteins A, B, C and E.¹⁰

Active amino acid transport via micro-villous and basal membranes of the trophoblast together with placental synthesis of some non-essential amino acids result in higher amino acid concentrations in fetal than in maternal serum.^{11,12}

Studies in preeclampsia and type 1 diabetic mothers have reported a significant increase in most amino acids concentrations in both mother and fetus.^{13,14} In women with preeclampsia several amino acids demonstrated significant inverse correlations with fetal head circumference.¹⁴ In diabetic women a significant changes in fetal/maternal amino acid concentration differences were also shown, suggesting alteration in placental amino acids exchange and/or fetal/placental amino acids metabolism.¹³

However, no data are available on the relationship between amino acid concentrations in both newborn infant and maternal serum in pregnancies associated with maternal obesity.

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The objective of this investigation was to study maternal cholesterol and lipoprotein metabolism and maternal and fetal amino acid concentrations from pregnancies associated with maternal obesity in two groups of obese women who were born with normal and low birth weight themselves. We hypothesize to find differences in pregnancy outcome and serum amino acids concentrations in both mother and fetus when maternal intrauterine development was normal or complicated by growth restriction. We also hypothesize there exists a relationship between apolipoprotein and maternal serum amino acid concentrations.

Materials and methods

Subjects

A total of 118 nulliparous subjects were recruited (2007–2012) and gave their informed consent at admission to labor and delivery at Mother and Child Research Institute (Yekaterinburg, Russia) as a part of ongoing investigation of maternal obesity approved by the institutional ethics committee.

Twenty-eight healthy, normal weight $(BMI < 25 \text{ kg/m}^2)$ with no significant past medical history women were included in the control group. Forty-six women with obesity $(BMI > 30 \text{ kg/m}^2)$ and normal birth weight were included in Ob-AGA group. Forty-four pregnant women with obesity $(BMI > 30 \text{ kg/m}^2)$ who themselves were small for gestational age (SGA) at birth were included in Ob-SGA group (Table 1). All women were non-smokers. The information on maternal birth weight was obtained from medical documentation related to birth certificate kept by the parents of pregnant women. Small for gestational age women were defined as those who were less than the 10th percentile for birth weight corrected for gestational age and sex.¹⁵ The criteria for exclusion in the control, Ob-AGA and Ob-SGA groups were hypertension, proteinuria and hyperuricemia. Hypertension was defined as a blood pressure more than 140/90 mmHg. Proteinuria was defined as >300 mg per 24 h urine collection. Hyperuricemia was defined as >5.5 mg/dl at term.

Fetal gestational age was calculated from the last menstrual period and confirmed by ultrasound before 20 weeks of gestation. For the neonates, gestational age at delivery, birth weight and length, sex and placenta weight were recorded.

Blood samples

Blood samples were collected at the time of delivery from maternal brachial vein after a 12-h overnight fasting as well as from umbilical vein in tubes containing $1.0 \text{ mg/ml} \text{ Na}_2\text{EDTA}$.

Maternal and cord-blood serum samples were allowed to stand at room temperature for 1 h, centrifuged at 2000 g for 20 min, aliquoted under sterile conditions and then stored at -80°C until they were assayed. For analysis plasma was quickly thawed and deproteinized with a solution of 10% sulfosalicylic acid with norleucine added as an internal standard and buffered with lithium hydroxide to pH 2.2. Samples were centrifuged at

Table 1. Clinical characteristics of the mothers and infants involved in the study

Parameter	Normal pregnancy $(n = 28)$	Ob-AGA group $(n = 46)$	Ob-SGA group $(n = 44)$
Mothers	· · ·	· · ·	
Age at delivery (years)	29.8 ± 6.3	31.1±5.4	32.2 ± 5.6
$BMI (kg/m^2)$	22.1 ± 1.3	33.4 ± 1.6^{a}	34.1 ± 1.8^{a}
Birth weight (g)	3437 ± 344	3276 ± 234	$2523 \pm 105^{a,b}$
Neonates			
Gestational age (weeks)	38.4 ± 0.2	38.3 ± 0.3	38.1 ± 0.4
Birth weight (g)	3290 ± 60	3276 ± 234	$2632 \pm 102^{a,b}$
Placental weight (g)	400.5 ± 89.1	495.5 ± 96.2	420.0 ± 67.2
Fetal weight/	8.22 ± 0.65	6.62 ± 2.37	6.26 ± 1.53
placental weight			
ratio			
Sex			
Male	16	21	24
Female	12	25	20

^aP < 0.05 compared with normal pregnancy values ^bP < 0.05 compared with Ob-AGA group.

14,000 rpm for 10 min and the supernatant fraction was filtered throughout a Millipore (Millipore Corp, Bedford, MA, USA) filter. Plasma amino acid concentrations were determined using automatic amino acid analyser AAA-T339M (Mikrotechna Praha, Czech Republic) according to the user's manual.

Fetal-maternal amino acid gradient was calculated as a ratio between fetal and maternal concentration of certain amino acid and was used for the assessment of placental concentration function. Maternal glucose, cholesterol and other lipid and lipoproteins fractions were determined by automated clinical analyzer Sapphire 400 (Tokyo Boeki Ltd, Japan).

Statistical analysis

Means and standard errors are reported. After confirming normality of dependent variables by the Kolmogorov–Smirnov test, we analyzed the data. A *t*-test was used to compare means. ANOVA, followed by Tukey's test, was used to compare three means. Correlation coefficients between ApoA, ApoB concentrations and amino acid concentrations were calculated by Pearson's method. The STATISTICA 10.0 (StatSoft) package was used. Significance was accepted at P < 0.05.

Results

The characteristics of the subjects are shown in Table 1. The mothers of all groups were matched for the age. There were significant differences for body mass index (BMI) between mothers of the control and both obesity groups. The mean birth weight of Ob-SGA group of mothers was significantly lower than that of the control and Ob-AGA groups of mothers.

The mean birth weight of infants of the Ob-AGA group was not essentially different from that of infants in the control group. The infants of Ob-SGA group mothers were on average smaller at birth than the infants from mothers of the control and Ob-AGA groups. There were no significant changes in placental weight and fetal weight/placental weight ratios between all groups of women.

Maternal cholesterol and lipoprotein concentrations

The main serum total cholesterol concentration was significantly higher in mothers of Ob-SGA group than in both the control and Ob-AGA group (Table 2). Triglyceride concentration was higher in Ob-SGA group than in the control group. High-density lipoprotein (HDL) cholesterol was significantly less in both Ob-AGA and Ob-SGA groups than in the control group of mothers. However, low-density lipoprotein (LDL) cholesterol was increased only in Ob-SGA group, apolipoprotein AI was also significantly increased in Ob-SGA group in comparison with Ob-AGA and the control groups. The most dramatic difference between groups was shown in total cholesterol/HDL ratio, which was almost two-fold higher in Ob-SGA group than in the control. This ratio was also significantly increased in Ob-AGA group comparing to the control group. There were no significant differences between fasting glucose concentrations in all groups of mothers.

Maternal amino acids in obese women

The mean concentration of most maternal amino acids was lower in both groups of obese women. Results for all amino acids required for protein synthesis and their summed total are shown in Table 3. Nine essential and five non-essential amino acids concentrations (except tyrosine) were significantly lower in Ob-AGA and Ob-SGA groups than in the control group mothers. However, the decrease in amino acid concentrations was more prominent in Ob-SGA group. The concentration of lysine in Ob-SGA group was two times lower than in the control group. Lysine concentration in Ob-AGA main group was only 1.3 times lower than in the control.

The mean total amino acid concentration for women of Ob-SGA group (525.1 \pm 64.0 μ M/l) was significantly decreased compared with women of Ob-AGA (845.6 \pm 83.8 μ M/l) and the control (1547.2 \pm 120.3 μ M/l) groups. Also there was a significant difference between total amino acid concentration in Ob-AGA and the control groups.

Table 3. Maternal plasma amino acid concentrations $(\mu M/l)$ in control, Ob-AGA and Ob-SGA groups

Amino acid	Normal pregnancy $(n = 28)$	Ob-AGA group $(n = 46)$	Ob-SGA group $(n = 44)$
Essential	()	(,, _, _,	(1)
Тгр	66.9 ± 2.9	18.6 ± 3.1^{a}	18.0 ± 5.5^{a}
Lvs	146.4 ± 18.5	109.8 ± 10.2	$72.7 \pm 6.2^{a,b}$
His	92.2 ± 5.1	65.0 ± 4.2^{a}	$32.1 \pm 3.6^{a,b}$
Thr	182.2 ± 12.1	118.3 ± 17.3^{a}	108.8 ± 11.5^{a}
Val	161.4 ± 7.5	54.1 ± 6.2^{a}	58.1 ± 4.9^{a}
Met	24.5 ± 6.5	6.4 ± 1.2^{a}	6.5 ± 0.8^{a}
Leu	118.4 ± 9.5	58.6 ± 6.0^{a}	$36.5 \pm 5.3^{a,b}$
Ile	48.9 ± 6.4	28.6 ± 7.3^{a}	13.8 ± 5.5^{a}
Phe	83.4 ± 7.2	35.8 ± 8.1^{a}	23.5 ± 7.4^{a}
Non-esse	ential		
Arg	82.3 ± 4.7	65.1 ± 3.6^{a}	$27.2 \pm 3.1^{a,b}$
Ser	83.9 ± 5.7	54.5 ± 4.5^{a}	$39.4 \pm 3.6^{a,b}$
Ala	245.8 ± 29.4	179.4 ± 15.6^{a}	$73.7 \pm 6.4^{a,b}$
Glu	56.8 ± 6.8	48.3 ± 5.4	$27.9 \pm 4.9^{ m a,b}$
Tyr	35.6 ± 4.1	32.7 ± 3.6	24.2 ± 2.8
Gly	118.5 ± 6.9	105.4 ± 6.5	$88.9 \pm 9.5^{a,b}$
Total	1547.2 ± 120.3	845.6 ± 83.8^{a}	$525.1\pm64.0^{a,b}$

 $^{a}P < 0.05$ compared with normal pregnancy values.

 $^{b}P < 0.05$ compared with Ob-AGA group.

Table 2. Mean fasting lipid, lipoprotein and glucose concentrations in pregnant women of control and two obese groups

Parameter	Normal pregnancy $(n = 28)$	Ob-AGA group ($n = 46$)	Ob-SGA group $(n = 44)$
Total cholesterol (mM/l)	5.39 ± 0.41	$5.04 \pm 0.35^{\circ}$	$6.94 \pm 0.46^{a,b}$
Triglyceride (mM/l)	2.05 ± 0.32	2.65 ± 0.28	3.34 ± 0.46^{a}
HDL cholesterol (mM/l)	1.81 ± 0.26	1.26 ± 0.19^{a}	1.04 ± 0.14^{a}
LDL cholesterol (mM/l)	3.03 ± 0.28	3.14 ± 0.19	$3.82 \pm 0.14^{a,b}$
ApoAI (g/l)	1.85 ± 0.29	2.24 ± 0.40	$3.64 \pm 0.68^{a,b}$
ApoAII (g/l)	0.59 ± 0.09	0.69 ± 0.18	0.79 ± 0.19
ApoB (g/l)	1.18 ± 0.20	2.01 ± 0.32	2.32 ± 0.13^{a}
Total cholesterol/HDL	2.97 ± 0.18	4.01 ± 0.58^{a}	$6.57 \pm 0.96^{a,b}$
Glucose (mM/l)	3.68 ± 0.45	3.79 ± 0.43	4.23 ± 0.58

 $^{a}P < 0.05$ compared with normal pregnancy values.

 $^{b}P < 0.05$ compared with Ob-AGA group.

Relationship between amino acid concentrations and ApoA, ApoB in maternal plasma

As shown in Table 4 there are negative correlations between ApoA and threonine in the Ob-SGA group, between ApoA and valine, methionine, leucine, arginine in all three groups of women and between ApoA and serine in the Ob-AGA and Ob-SGA groups. Table 5 demonstrates inverse correlation between ApoB and lysine and hystidine in Ob-AGA and Ob-SGA groups and between ApoB and following amino acids: valine, arginine, serine, alanine and glycine in all three groups of women.

Cord blood amino acid concentrations

Obese pregnancy groups had lower umbilical venous plasma amino acid concentrations (except glutamine, tyrosine and glycine for Ob-AGA and glutamine for Ob-SGA groups) than the control pregnancy group (Table 6).

Table 4. Correlation coefficients and P-values between ApoA and amino acid concentrations in maternal plasma

	Normal pregnar	ncy (n = 28)	Ob-AGA grou	p ($n = 46$)	Ob-SGA grou	p ($n = 44$)
Amino acid	Coefficient	P-value	Coefficient	<i>P</i> -value	Coefficient	<i>P</i> -value
Essential						
Trp	0.159	0.458	0.241	0.145	-0.137	0.210
Lys	-0.314	0.135	-0.362	0.084	-0.255	0.230
His	-0.191	0.372	-0.154	0.631	-0.345	0.029
Thr	-0.331	0.036*	-0.296	0.041	-0.446	0.021*
Val	-0.407	0.047*	-0.395	0.045*	-0.497	0.015*
Met	-0.395	0.048*	-0.496	0.018*	-0.398	0.031*
Leu	-0.397	0.041*	-0.510	0.008*	-0.570	0.006*
Ile	-0.417	0.039*	-0.386	0.048*	-0.510	0.019*
Phe	-0.028	0.850	-0.115	0.501	-0.361	0.085
Non-essential						
Arg	-0.446	0.029*	-0.339	0.012*	-0.467	0.028*
Ser	-0.312	0.176	-0.481	0.021*	-0.448	0.031*
Ala	-0.510	0.009*	-0.501	0.008^{*}	-0.495	0.021*
Glu	-0.345	0.291	-0.316	0.187	-0.215	0.101
Tyr	-0.287	0.174	-0.159	0.451	-0.203	0.341
Gly	0.251	0.219	-0.287	0.174	-0.335	0.109

**P*<0.05.

Table 5. Correlation coefficients and P-values between ApoB and amino acid concentrations in maternal plasma

	Normal pregnat	ncy (n = 28)	Ob-AGA grou	p ($n = 46$)	Ob-SGA grou	p ($n = 44$)
Amino acid	Coefficient	<i>P</i> -value	Coefficient	<i>P</i> -value	Coefficient	<i>P</i> -value
Essential						
Trp	0.260	0.218	-0.162	0.312	-0.135	0.134
Lys	-0.364	0.079	-0.415	0.021*	-0.475	0.019*
His	-0.181	0.367	-0.445	0.029*	-0.396	0.041*
Thr	-0.221	0.101	-0.289	0.175	-0.334	0.106
Val	-0.508	0.010*	-0.484	0.032*	-0.401	0.045*
Met	-0.203	0.342	-0.160	0.605	-0.204	0.319
Leu	-0.202	0.296	-0.109	0.675	-0.221	0.321
Ile	-0.054	0.810	-0.261	0.270	-0.250	0.213
Phe	-0.318	0.091	-0.396	0.012	-0.287	0.201
Non-essential						
Arg	-0.452	0.038*	-0.519	0.018*	-0.578	0.019*
Ser	-0.496	0.034*	-0.528	0.027*	-0.594	0.014^{*}
Ala	-0.498	0.030*	-0.596	0.018*	-0.481	0.043*
Glu	-0.315	0.287	-0.216	0.087	-0.267	0.121
Tyr	-0.137	0.186	-0.396	0.076	-0.328	0.065
Gly	0.598	0.021*	-0.479	0.031*	-0.489	0.016*

**P* < 0.05.

Amino acid	Normal pregnancy $(n = 28)$	Ob-AGA group $(n = 46)$	Ob-SGA group $(n = 44)$
Essential			
Trp	88.4 ± 9.3	21.7 ± 3.7^{a}	14.6 ± 3.4^{a}
Lys	372.7 ± 25.8	145.1 ± 17.4^{a}	$107.1 \pm 12.5^{a,b}$
His	169.5 ± 9.6	74.7 ± 6.5^{a}	$38.4 \pm 3.7^{a,b}$
Thr	268.4 ± 32.4	169.7 ± 24.7^{a}	$114.1 \pm 19.5^{a,b}$
Val	218.7 ± 34.1	60.8 ± 4.7^{a}	59.7 ± 4.5^{a}
Met	30.6 ± 2.4	7.9 ± 2.1^{a}	$12.2 \pm 2.9^{a,b}$
Leu	123.5 ± 18.9	66.9 ± 8.1^{a}	48.1 ± 6.2^{a}
Ile	67.4 ± 6.1	32.6 ± 5.2^{a}	$20.7 \pm 4.6^{a,b}$
Phe	88.4 ± 8.7	39.7 ± 6.4^{a}	39.8 ± 3.6^{a}
Non-esse	ential		
Arg	97.7 ± 11.7	34.2 ± 3.8^{a}	$26.3\pm2.4^{\rm a}$
Ser	142.6 ± 31.5	59.8 ± 7.9^{a}	54.2 ± 6.2^{a}
Ala	320.1 ± 38.6	163.2 ± 12.5^{a}	$102.4 \pm 8.4^{a,b}$
Glu	28.9 ± 3.7	23.4 ± 1.5	23.1 ± 5.0
Tyr	62.6 ± 8.7	58.2 ± 6.1	$32.2 \pm 5.5^{a,b}$
Gly	241.7 ± 48.4	164.3 ± 27.5	123.2 ± 18.6^{a}
Total	2321.3 ± 289.9	1122.2 ± 138.1^{a}	816.1 ± 107.0^{a}

Table 6. Newborn infants plasma amino acid concentrations (μ M/l) in control, Ob-AGA and Ob-SGA groups

 $^{a}P < 0.05$ compared with normal pregnancy values.

 $^{b}P < 0.05$ compared with Ob-AGA group.

Table 7. Fetal-maternal amino acid gradients for control andobesity groups

Amino acid	Normal pregnancy $(n = 28)$	Ob-AGA group $(n = 46)$	Ob-SGA group $(n = 44)$
E		· · ·	· · ·
Essential			
Trp	1.31 ± 0.17	1.16 ± 0.10	0.81 ± 0.07^{a}
Lys	2.54 ± 0.24	1.31 ± 0.18^{a}	1.57 ± 0.11^{a}
His	1.83 ± 0.18	1.49 ± 0.09^{a}	1.19 ± 0.12^{a}
Thr	1.47 ± 0.17	1.43 ± 0.19	1.05 ± 0.06
Val	1.35 ± 0.21	1.11 ± 0.14	0.91 ± 0.06^{a}
Met	1.22 ± 0.14	1.13 ± 0.15	1.32 ± 0.09
Leu	1.04 ± 0.08	1.15 ± 0.07	1.31 ± 0.10
Ile	1.36 ± 0.14	1.15 ± 0.18	1.49 ± 0.16
Phe	1.06 ± 0.05	1.12 ± 0.16	1.70 ± 0.21
Non-esse	ential		
Arg	1.95 ± 0.20	0.52 ± 0.06^{a}	$0.97 \pm 0.04^{ m a,b}$
Ser	1.69 ± 0.18	1.10 ± 0.14^{a}	1.38 ± 0.11
Ala	1.30 ± 0.09	0.91 ± 0.07^{a}	1.39 ± 1.39
Glu	0.50 ± 0.04	0.48 ± 0.03	$0.82\pm0.09^{\rm a}$
Tyr	1.77 ± 0.12	1.75 ± 0.13	1.32 ± 0.14
Gly	2.04 ± 0.16	1.76 ± 0.14^{a}	$1.30 \pm 0.09^{a,b}$

 $^{a}P < 0.05$ compared with normal pregnancy values.

 $^{b}P < 0.05$ compared with Ob-AGA group.

In normal pregnancy umbilical venous plasma concentrations for most essential and non-essential amino acids (except leucine and glutamine) were significantly higher than maternal concentrations (high fetal-maternal amino acids gradient) (Table 7). The fetalmaternal amino acid gradients were lowest in Ob-SGA group. However, for several amino acids (Met, Leu, Ile, Phe) these gradients were higher than in Ob-AGA and the control groups.

Discussion

This study shows a significant increase in serum lipids which is more prominent in the Ob-SGA group. Women of this group had serum concentration of total cholesterol that would be considered as an increased risk of cardiovascular disease in nonpregnant women. We have also shown that the concentrations of LDL cholesterol and their corresponding apolipoproteins B and AI are increased in women of Ob-SGA group. The increase in total, LDL cholesterol and triglyceride has been demonstrated during normal pregnancy.^{16,17} The mechanism whereby pregnancy induces hyperlipidemia has not been fully elucidated. The positive correlation between changes in the lipid and lipoprotein concentration and pregnancy hormones (estradiol, progesterone and human placental lactogen) has been demonstrated.¹⁸

Although the significance of these changes in plasma lipids and lipoproteins is uncertain, they are likely to relate to the maintenance of energetic fuel to the fetus.¹⁹

Estrogens could be responsible for most of the changes in lipoprotein metabolism during pregnancy, but its effects are complemented and opposed by the other pregnancy hormones (progesterone) and by increasing insulin resistance in late pregnancy. Our study demonstrates that lipid and lipoprotein concentration in obese pregnant women who themselves were born SGA could be significantly increased in comparison with the control women and women with diet-induced obesity. We suggest that potentially the Ob-SGA group could be atherogenic. The changes in lipid profile observed in women of Ob-SGA group may be of potential importance for women's long-term health because elevated serum triglycerides are independent risk factors for coronary heart disease in women.²⁰

The complex nature of apolipoproteins synthesis and metabolism has been demonstrated in many studies based upon the use of labeled amino acids.^{21,22} In this study we assessed the relationship between ApoA, ApoB and several amino acids concentrations in maternal plasma. Negative correlations (correlation coefficients from -0.3 to -0.5) were seen between these apolipoproteins and several amino acids concentrations. Negative correlation between ApoA and threonine concentration was demonstrated only in Ob-SGA group and between ApoA and serine concentration in both groups of obese mothers. Inverse correlations between ApoB and lysine, hystidine concentrations were also seen only in obese women. Earlier the inverse association was demonstrated between intakes of alanine, isoleucine, methionine, serine, tryptophan, tyrosine and ApoA/ApoB ratio among female adolescents.²³ One can speculate that the increase of the amino acids contribution to the synthesis of apolipoproteins could diminish the role of such amino acids as an energetic fuel to the fetus.

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It was shown earlier, that women who were SGA at birth are at increased risk of developing hypertension during pregnancy.²⁴ Also it was shown that maternal hypercholesterolemia dramatically raised the cumulative area of intimal lipid accumulation and the area of the largest lesion per fetal aortic section, increasing offspring risk of future cardiovascular disease.²⁵

Our results support the Barker theory that malnutrition and impaired fetal growth are associated with an increased risk of developing a variety of risk factors for cardiovascular disease, including dyslipidemia during adulthood.²⁶

Amino acids

Amino acids along with glucose and lactate represent major nutrients used by fetus both for protein synthesis and oxidation.⁸ It is demonstrated that amino acid concentrations are higher in the fetal than in maternal body. Number of studies has provided evidence for significant participation of placenta in amino acid metabolism.²⁷

Amino acids uptake across the syncytiotrophoblast membranes is mediated through active transporters and exchangers. Active accumulative transporters increase intracellular amino acid concentrations by transferring these substances against their concentration gradient, usually by co-transporting extracellular sodium.²⁸ Exchangers are able to modify amino acid content by exchanging amino acids between intracellular and extracellular compartments.

The placenta not only concentrates amino acids in the fetal compartment but is also involved in synthesis of some nonessential amino acids. As recently demonstrated, some external factors may regulate the activity of amino acid transporters (such as oxygenation, insulin, leptin).²⁹

Fetal intrauterine growth is the result of interaction between maternal–placental nutrient supply and genetically determined growth potential.⁸ Some factors may influence fetal nutritional supply: maternal nutrition, uteroplacental circulation, and placental transfer capacity.³⁰ We demonstrated that newborn infants of Ob-SGA group women were born SGA. Moreover, in this group umbilical concentrations for most amino acids were significantly lower than in the control and even in Ob-AGA group. Earlier it was shown that concentration of most amino acids is significantly decreased both in the umbilical artery and vein of intrauterine growth restriction (IUGR) pregnancies when it compared to normally grown babies.^{31,32} However, in IUGR maternal concentrations of the most essential amino acids were significantly higher than in pregnancies with appropriate for gestational age fetuses.³¹

In IUGR pregnancies, increasing the maternal concentration of amino acids leads to an increased umbilical uptake of some of the amino acids to the fetus. However, the changes in the uptake of the essential amino acids such as lysine, histidine, threonine, valine, and phenylalanine suggest the presence of competition for the same transporter that might diminish the transport.³³ Studies in human pregnancies demonstrate that during constant infusion of L-[1-13C]-leucine, the fetal–maternal leucine gradient decreases in IUGR in parallel with clinical severity.³³ It was

suggested than not only the transplacental transport of leucine is impaired but also a possible increased protein catabolism in IUGR pregnancies.³⁵ The injection as a bolus of two essential amino acids, leucine and phenylalanine demonstrated in IUGR pregnancies decreased fetal–maternal gradient in comparison with AGA pregnancies, whereas no differences were presented for the nonessential amino acids glycine and proline.³⁴

We have demonstrated that Ob-SGA group had the lowest fetal-maternal amino acid gradients for most amino acids that could reflect the impaired concentration/synthetic function of the placenta. However, these gradients for Met, Leu, Ile, Phe were higher in Ob-SGA group than in other groups of women. It could be the result of changing the balance between placental amino acid transporting systems in such type of obesity. Many types of these systems have been identified in the placenta.⁸ Each transporter is highly stereospecific, but different transporting systems have overlapping substrate specificity, with possible compensation of one system by another.⁸

It is interesting that we demonstrated increase in fetalmaternal gradient for leucine and phenylalanine in OB-SGA group in contrary to the results shown by Paolini et al.³⁴ One can speculate that changes in placental transporting function in obese mothers born themselves SGA significantly differ from corresponding placental changes in IUGR.

The fetal growth and metabolism are adaptive processes and programmed by the intrauterine nutrition and environment.³⁵ Furthermore, as suggested earlier, the placenta could play a role of a nutrient sensor.³⁶ If the placenta senses the fetal environment with low nutrient levels, it increases its transport activity to support normal fetal growth. On the other hand, if there is an insufficient nutrient supply at the maternal site, the placenta may diminish its transport activity, adapting fetal growth to a lower level, in order to reduce the postnatal demand.³¹

In our study, decreased fetal-maternal amino acid gradients for most amino acids observed in Ob-SGA group suggest that placental amino acid exchange and/or fetal-placental metabolism are altered in obese women who were born SGA. We can also suggest that hypercholesterolemia in Ob-SGA women could affect the fetal amino acid metabolism due to the reciprocal relationship between fetal energy supply and placental amino acid transport. One can speculate that significant changes in lipids and apolipoproteins metabolism during the intrauterine development of Ob-SGA women affected future placental function in a specific manner that resulted in the development of low birth weight infants.

Limitation

The limitation of this research project is that authors have no complete information on the cause of maternal SGA women of Ob-SGA group.

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References

- Drake A, Reinolds R. Impact of maternal obesity on offspring obesity and cardiometabolic disease risk. *Reproduction*. 2010; 140, 387–398.
- 2. Blackmore HL, Ozanne SE. Maternal diet-induced obesity and offspring cardiovascular health. *J Develop Origins Health Dis.* 2013; 4, 338–347.
- Stettler N, Zemel BS, Kumanika S, Stallings VA. Infant weight gain and childhood overweight status in a multicenter cohort study. *Pediatrics*. 2002; 109, 194–199.
- Dietz WH. Overweight in childhood and adolescence. N Engl J Med. 2004; 350, 855–857.
- Barker DJP, Osmond C, Forsén T, Kajantie E, Eriksson JG. Trajectories of growth among children who have coronary events as adults. *N Engl J Med.* 2005; 353, 1802–1809.
- Hales CN, Barker DJP, Clark PMS, *et al.* Fetal and infant growth and impaired glucose tolerance at age 64. *BMJ*. 1991; 303, 1019–1022.
- Lithell UB, Leon DA. Relation of size at birth to non-insulin dependent diabetes and insulin concentrations in men aged 50–60 years. *BMJ*. 1996; 312, 406–410.
- 8. Battaglia FC, Regnault TRH. Placental transport and metabolism of amino acids. *Placenta*. 2001; 22, 145–161.
- Parhofer KG, Barrett HR, Bier DM, Schonfeld G. Determination of kinetic parameters of apolipoprotein B metabolism using amino acids labeled with stable isotopes. *J Lipid Res.* 1991; 32, 1311–1323.
- Lichtenstein AH, Cohn JS, Hachey DL, *et al.* Comparison of deuterated leucine, valine, and lysine in the measurement of human apolipoprotein A-I and B-100 kinetics. *J Lipid Res.* 1990; 31, 1693–1701.
- Young M, Prenton MA. Maternal and fetal plasma amino acid concentrations during gestation and in retarded fetal growth. *J Obstet Gynaec Br Commonw.* 1969; 76, 333–344.
- Johnson LW, Smith CH. Neutral amino acid transport systems of microvillous membrane of human placenta. *Am J Physiol.* 1988; 254, C773–C780.
- Kalkhoff RK, Kandaraki E, Morrow PG, *et al.* Relationship between neonatal birth weight and maternal plasma amino acids profiles and lean and obese nondiabetic women and type 1 diabetic pregnant women. *Metabolism.* 1988; 37, 234–239.
- Evans RW, Powers RW, Ness RB, *et al.* Maternal and fetal amino acid concentrations and fetal outcomes during pre-eclampsia. *Reproduction.* 2003; 125, 785–790.
- 15. WHO child growth standards based on length/height, weight and age. *Acta Paediatrica*. 2006; 95(Suppl. S459), 76–85.
- Ordovas JM, Pocovi M, Grande F. Plasma lipids and cholesterol esterification during pregnancy. *Obstet Gynecol.* 1984; 63, 20–25.
- Piechota W, Staslewski A. Reference ranges of lipids and apolipoproteins in pregnancy. *Eur J Obstet Gynecol Reproduct Biol.* 1992; 45, 27–35.
- Desoye G, Schweditsch MO, Pfeiffer KP, Zechner L, Kostmer GM. Correlation of hormones with lipid and lipoprotein levels during normal pregnancy and postpartum. *J Clin Endocrinol Metab.* 1987; 64, 704–712.
- Martin U, Davies C, Hayavi S, Hartland A, Dunne F. Is normal pregnancy atherogenic? *Clin Sci.* 1999; 96, 421–425.

- Hachey DL. Benefits and risks of modifying maternal fat intake in pregnancy and lactation. *Am J Clin Nutr.* 1994; 59, 454S–464S.
- Patterson BW, Hachey DL, Cook GL, *et al.* Metabolic kinetics of apolipoproteins C using a stable isotope amino acid tracer. *Arteriosclerosis.* 1989; 9, 757a–758a.
- 22. Cohn JS, Wagner DA, Cohn SD, Millar JS, Schaefer EJ. Measurement of very low density and low density lipoprotein apolipoprotein (Apo) B-100 and high density lipoprotein ApoA-1 production in human subjects using deuterated leucine (effect of fasting and feeding). *J Clin Invest*. 1990; 85, 804–811.
- Bel-Serrat S, Mouratidou T, Huybrechts I, *et al.* The role of dietary fat on the association between dietary amino acids and serum lipid profile in European adolescents participating in the HELENA Study. *Eur J Clin Res.* 2014; 68, 464–473.
- Klebanoff MF, Secher NJ, Mednick BR, Schulsinger S. Maternal size at birth and the development of hypertension during pregnancy. A test of the Barker hypothesis. *Arch Intern Med.* 1999; 159, 1607–1612.
- Napoli C, D'Armiento FP, Mancini FP, *et al.* Fatty streak formation occurs in human fetal aortas and is greatly enhanced by maternal hypercholesterolemia. Intimal accumulation of low density lipoprotein and its oxidation precede monocyte recruitment into early atherosclerotic lesions. *J Clin Invest.* 1997; 100, 2680–2690.
- Barker DJP. Maternal nutrition, fetal nutrition, and disease in later life. *Nutrition*. 1997; 13, 807–813.
- Cetin I. Amino acid interconversions in the fetal-placental unit: the animal model and human studies in vivo. *Pediatr Res.* 2001; 49, 148–154.
- Broer S. Adaptation of plasma membrane amino acid transport mechanisms to physiological demands. *Pflugers Archiv Eur J Physiol.* 2002; 444, 457–466.
- Desforges M, Greenwood SL, Glazier JD, Westwood M, Sibley CP. The contribution of SNAT1 to system A amino acid transporter activity in human placental trophoblast. *Biochem Biophys Res Commun.* 2010; 398, 130–134.
- Jansson T, Ylven K, Wennergren M, Powell TL. Glucose transport and system A activity in syncytiotrophoblast microvillous and basal plasma membranes in intrauterine growth restriction. *Placenta*. 2002; 23, 392–399.
- Economides DL, Nicolaides KH, Gahl WA, Bernardini I, Evans MI. Plasma amino acids in appropriate- and small-for-gestationalage fetuses. *Amer J Obstet Gynecol.* 1989; 161, 1219–1227.
- Cetin I, Corbetta C, Sereni L, *et al.* Umbilical amino acid concentrations in normal and growth-retarded fetuses sampled in utero by cordocentesis. *Amer J Obstet Gynecol.* 1990; 162, 253–261.
- Marconi AM, Paolini CL, Stramare L. Steady state maternal-fetal leucine enrichment in normal and intrauterine growth-restricted pregnancies. *Pediatr Res.* 1999; 46, 114–119.
- Paolini CL, Marconi AM, Ronzoni S. Placental transport of leucine, phelalanine, glycine, and proline in intrauterine growthrestricted pregnancies. *J Clin Edocrinol Metab.* 2001; 86, 5427–5432.
- 35. Solomons NW. Developmental origins of health and disease: concepts, caveats, and consequences for public health nutrition. *Nutr Rev.* 2009; 67, S12–S16.
- Jansson T, Powell TL. Human placental transport in altered fetal growth: does the placenta function as a nutrient sensor? *Placenta*. 2006; 27, 91–97.